Supplementary Table S2. Study on *in vitro* production methods of *Gloriosa superba* L. via indirect organogenesis in various callus-derived explants[1].

Sr. No	Explant(s) of interest	The best treatment for callus induction	Final goal	Best treatment for achieving the final goal	Shoot initiation percentage (%)
1	Nodal segments	MS basal media + 4 mg/l 2,4-D + 5 mg/l KN + Media conditions: 20% coconut water, 0.8% (w/v) agar, pH of 5.8, 25±2°C, 16/8-hour light/dark	Indirect somatic embryogenesis	MS basal media + 10 mg/l CH + 5 mg/l kinetin + 4 mg/l 2,4-D Media conditions: 20% coconut water, 0.8% (w/v) agar, pH of 5.8, 25±2°C, 16/8-hour light/dark	-
2	Auxillary Buds	MS basal media + 4.52 µM 2,4-D + 13.28 µM BAP Media conditions: 20% coconut water, 0.8% (w/v) agar, pH of 5.8, 25±2°C, 16/8-hour light/dark	Shoot morphogenesis	MS basal media + 4.50 µM 2,4-D + 17.80 µM BAP Media conditions: 20% coconut water, 0.8% (w/v) agar, pH of 5.8, 25±2°C, 16/8-hour light/dark	-
3	Corm Buds (4 – 6 weeks old in vitro seedlings)	MS basal media + 10 mg/l 2,4-D + 5 mg/l IAA Media conditions: 0.8% agar + 30% sucrose	Shoot morphogenesis	MS basal media + 1.5 mg/l BAP + 0.2 mg/l NAA + 15% coconut water + 2 g/l acetone Media conditions: 0.8% agar + 30% sucrose	-
4	Root	MS basal media + 2 mg/l 2,4D + 1 mg/l IAA + 0.75 mg/l NAA Media conditions: 0.8% agar + 30% sucrose	Shoot morphogenesis	MS basal media + 3 mg/l BAP + 1 mg/l IBA + IPA 0.75 mg/l Media conditions: 0.8% agar + 30% sucrose	90.60%

(-) means data was not documented in the study

REFERENCE

[1] Gurung R, Sharma S, Sharma S, Sharma V. 2021. *Gloriosa superba*: Its properties and *in vitro* production methods. *International Journal of Botany Studies* 6(3):74–77.