

Supporting information

Relative contributions of denitrification and anammox to nitrogen removal in riverine wetlands across China

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Determination of nitrogen removal potential

The denitrification and anammox rates of channel sediments, riparian rhizosphere soils, and depth-selected riparian bulk soils (0–20, 60–80, and 160–180 cm in depth) were quantified using the ¹⁵N isotope pairing method [1,2]. Briefly, three 5 g replicates of homogenized sediment or soil samples were weighed into 12-mL cylindrical glass vials containing helium-purged distilled water [3]. Subsequently, all vials were pre-incubated in a horizontal shaker (180 rpm) at 24°C for 3 days to remove the background NO₃⁻ and NO₂⁻, as well as residual oxygen [4]. All vials were then divided into three groups and injected with 100 μL helium-purged stock solution: ¹⁵NH₄Cl (98 at.% ¹⁵N), ¹⁵NH₄Cl + K¹⁴NO₃, or ¹⁴NH₄Cl + K¹⁵NO₃ (99.99

at.% ^{15}N). This resulted in a final total N concentration of 100 μmol in each vial ^[5]. Additions of $^{15}\text{NH}_4^+$ solutions were used to verify that background NO_3^- -N and NO_2^- -N have been consumed, while additions of $^{15}\text{NH}_4^+$ -N + $^{14}\text{NO}_3^-$ -N solutions were used to confirm the presence of anammox process. Additions of $^{14}\text{NH}_4^+$ -N + $^{15}\text{NO}_3^-$ -N solutions were used to determine denitrification and anammox rates. The incubation was terminated at five-time intervals (0, 1, 2, 4, and 8 h) by injecting 200 μL of 7 M ZnCl_2 solution. The isotopic abundance of $^{28}\text{N}_2$, $^{29}\text{N}_2$, and $^{30}\text{N}_2$ in the glass vials were measured using isotope ratio mass spectrometry (Gasbench II and Delta V Advantage, Thermo Finnigan, Germany).

Analysis of abundance of denitrifying and anammox bacteria

DNA was extracted from replicate soil and sediment samples using a PowerSoil DNA Isolation Kit (MoBio Laboratories Inc., CA, USA), and DNA concentrations were determined using a NanoDrop 2000 Spectrophotometer (Thermo Fisher Scientific, MA, USA). The abundance (i.e., copy number) of *nirK*, *nirS*, and *hzsB* genes was measured in triplicate by using a Roche LightCycler480 real-time PCR system (Roche Diagnostics, Mannheim, Germany) with the fluorescent dye SYBR green qPCR method. The reduction of NO_2^- -N to nitric oxide (NO) is the rate-limiting step of denitrification, which is catalyzed by either a copper-containing enzyme encoded by the *nirK* gene or a cytochrome-cd1 enzyme encoded by the *nirS* gene^[6]. The *hzsB* gene encoded for hydrazine synthase β -subunit which catalyzes the production of hydrazine from NH_4^+ -N and NO_2^- -N during the anammox process ^[4]. Detailed procedures of qPCR were described previously^[7]. In brief, the abundance (i.e., copy number) of *nirS*, *nirK*, and *hzsB* genes was determined in triplicate using a Roche LightCycler480 real-time PCR System (version 1.5.0; Roche Diagnostics, Mannheim, Germany) with the fluorescent dye SYBR green quantitative PCR method. Primer sets of *nirSCd3aF/nirSR3cdR*, *nirKF1aCu/nirKR3Cu*, and *hzsB396F/hzsB742R* were applied for the *nirS*, *nirK*, and *hzsB* genes, respectively (Table S2). The 20- μL quantitative PCR mixture contained 10 μL of SyberGreen qPCR Master Mix (2 \times), 1 μL of primers (10 μM), and 2 μL of DNA template. The primers and qPCR protocol are listed in [Supplementary Table S2](#). Standard curves were constructed with serial plasmid dilutions of a known amount of plasmid DNA containing a fragment of the *nirS*, *nirK*, and *hzsB* genes.

Measurements of soil, sediment, and water physicochemical properties

Soil pH was measured at a soil-to-water ratio of 1:5 (v/v) using a pH meter (Hanna Instruments, Padova, Italy), whereas soil moisture content was determined gravimetrically by drying 20 g of fresh soil at 105°C

for 24 hours. The soil TN, TC, and TOC concentrations of air-dried soil samples were determined by an elemental analyzer. (Vario TOC cube, Hanau, Germany). Soil or sediment physicochemical properties, including moisture, pH, total N (STN), $\text{NH}_4^+\text{-N}$, $\text{NO}_3^-\text{-N}$, total carbon (STC) and organic carbon (SOC) were determined following published analytical methods [8,9]. Soil Fe^{2+} and AFe concentrations were measured using the phenanthroline spectrophotometric method [10]. The concentrations of water TN, $\text{NH}_4^+\text{-N}$, $\text{NO}_3^-\text{-N}$, TC, and TOC were measured in the laboratory, as described previously [11]. The concentrations of NH_4^+ and NO_3^- in water were analyzed by a continuous flow analyzer (EasyChem plus, Systea, Italy). The water TN, TC, and TOC concentrations of samples were determined by a Total Organic Carbon Analyzer Analyzer (Liquid). (Vario TOC cube, Hanau, Germany).

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